



**SWARNNIM**  
STARTUP & INNOVATION  
UNIVERSITY  
WHERE IDEAS COME ALIVE

INDIA'S FIRST UNIVERSITY FOR STARTUP

Ref: swarnnim/20/Letter/2025/76

Date: 06/09/2025

To

The Manager,

Spechrom Solutions.

Subject: Request for Issuance of Sanction Letters in the Name of Swarnnim Startup & Innovation University for Allocated Major Research Projects (AY 2019-2024)

Dear Sir,

We, Swarnnim Startup & Innovation University, would like to extend our sincere thanks for providing financial support and promote industry sponsored research throughout the year.

With reference to the above subject, we would like to inform you that Swarnnim Startup & Innovation University has received 15 research projects amounting to ₹1,89,50,000 during the academic year 2019-2024. Actually, these projects were given to the University but the letters were issued in the name of Constituent departments.

As per the recent query raised by NAAC, all sanction letters are required to be issued in the name of the University. Therefore, we kindly request you to re-issue the sanction letters/clarification letter in the name of Swarnnim Startup & Innovation University for the above-mentioned projects.

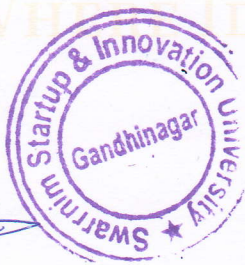
We sincerely appreciate your support and cooperation in this matter.

Your prompt response is highly appreciated considering the urgency of the matter,

Thanking you,

Yours faithfully,

Registrar



Swarnnim Startup & Innovation University

**Managed by G P Jain Charitable Trust**

**University Campus : Bhoyan Rathod, Opposite IFFCO, Near ONGC WSS, Adalaj Kalol Highway,  
Gandhinagar, Gujarat - 382422.**

**+91 95123 43333 | info@swarnnim.edu.in | www.swarnnim.edu.in**

Date: 12/09/2025

To,

Swarnnim Startup & Innovation University,

Bhoyan Rathod, Opposite IFFCO,

Near ONGC WSS, Adalaj Kalol Highway,

Gandhinagar, Gujarat - 382422.

**Subject:** Sanction Letter of Research Project

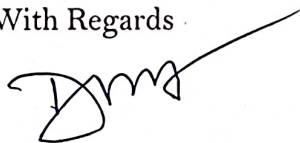
**Dear Researcher,**

I am pleased to inform you that the scrutiny committee has approved your research. The details of the Research Proposal and Sanctioned grant is as under.

Title of Research Proposal	Copper biosorption utilizing dehydrated Aspergillus biomass
Name of Principal investigator	Dr. Priti Mahla
Name of Co-investigators	Dhara Goti, Darshika Savalia, Narmatta Datta, Priyanka Shrimali, Mitesh Prajapati
Duration in months	12
Sanctioned Grant for Research Work	500000

It is further hereby informed that you will abide to follow all the guidelines/norms, terms, and conditions mentioned in the Research Scheme of company. The principal investigator may contact for any further assistance. All the best for great research work ahead.

With Regards





Date: 30/3/2020

To,

Swarnim Science collage,  
Bhoyan Rathod, Opposite IFFCO,  
Near ONGC WSS, Adalaj Kalol Highway,  
Gandhinagar, Gujarat - 382422.


Subject: Sanction Letter of Research Project

Dear Researcher,

I am pleased to inform you that the scrutiny committee has approved your research. The details of the Research Proposal and Sanctioned grant is as under.

Title of Research Proposal	Copper biosorption utilizing dehydrated Aspergillus biomass
Name of Principal investigator	Dr. Priti Mahla
Name of Co-investigators	Dhara Goti, Darshika Savalia, Narmatta Datta, Priyanka Shrimali, Mitesh Prajapati
Duration in months	12
Sanctioned Grant for Research Work	500000

It is further hereby informed that you will abide to follow all the guidelines/norms, terms, and conditions mentioned in the Research Scheme of company. The principal investigator may contact for any further assistance. All the best for great research work ahead.

  
With Regards



SWARANIM STARTUP AND INNOVATION UNIVERSITY

Period : 01-04-2018 To 06-04-2021

Cust.ReIn.No : 59701131

Account No : 9712631137

Currency : INR

Branch : AHMEDABAD - PALDI

Nominee Registered : N

35 WORLD BUSINESS HOUSE NR

PARIMAL GARDEN ELLISBRIDGE

AHMEDABAD-380006

GUJARAT,INDIA

Date	Narration	Chq/Ref No	Withdrawal (Dr)	Deposit(Cr)	Balance
16-12-2020	CLG TO VODAFONE MOBILE SERVICE CITI BANK	2075	685.00		18,897.95(Cr)
16-12-2020	CLG TO VODAFONE MOBILE SERVICE CITI BANK	2076	412.00		18,485.95(Cr)
16-12-2020	IB: FUND TRANSFER FROM 9712648883	IB		200,000.00	218,485.95(Cr)
16-12-2020	FUND TRANSFER TO JIGARKUMAR THAKKAR	2083	26,000.00		192,485.95(Cr)
16-12-2020	Sent NEFT KKBKH20351626231/ESSKSEE CONSULTANC	2082/000191804726	46,374.00		146,111.95(Cr)
17-12-2020	CLG TO SPICE HUB RESTRO CAFE 2042 INDUSIND BANK LIM		17,670.00		128,441.95(Cr)
18-12-2020	IB: FUND TRANSFER FROM 9712631168	IB		300,000.00	428,441.95(Cr)
18-12-2020	IB: FUND TRANSFER FROM 9712639331	IB		950,000.00	1,378,441.95(Cr)
18-12-2020	IB: FUND TRANSFER FROM 9712648876	IB		800,000.00	2,178,441.95(Cr)
18-12-2020	IB: FUND TRANSFER FROM 9712648883	IB		1,200,000.00	3,378,441.95(Cr)
18-12-2020	Sent RTGS KKBKR52020121800652499/GYAPR 4 ASAD JA	2085/00019209866	2,000,000.00		1,378,441.95(Cr)
18-12-2020	UPI/jaypatel28./035317346591/UPI	UPI-035317882494		2,500.00	1,380,941.95(Cr)
18-12-2020	Sent NEFT KKBKH20353655202/ANAND S MASHRUWALA	2090/000192100975	7,500.00		1,373,441.95(Cr)
18-12-2020	Sent NEFT KKBKH20353655541/SWASTIK INFRASTRUC	2084/000192100463	100,000.00		1,273,441.95(Cr)
19-12-2020	Sent NEFT KKBKH20354744400/WORLDWIDE DISTRIBU	2093/000192184908	5,000.00		1,268,441.95(Cr)
19-12-2020	FUND TRANSFER TO DEEP JAYESHKUMAR MEHTA	2092	4,510.00		1,263,931.95(Cr)
19-12-2020	RTGS CR-502000496721 SPECHROM SOLUTIONS	2088/000197287211		500,000.00	1,863,931.95(Cr)
19-12-2020	Sent RTGS KKBKR52020121900794921/GYAPR 7 ASAD JA	2086/00019226611	1,500,000.00		363,931.95(Cr)
19-12-2020	UPI/jay247k@oki/035421412409/Con vocation	UPI-035421567944		2,500.00	366,431.95(Cr)



## CERTIFICATE FROM THE INVESTIGATOR

**PROJECT Title :** Copper Biosorption Utilizing Dehydrated Aspergillus Biomass

**Name(s) of the PI's and Co-PI's:**

**Principal Investigator:** Dr. Priti Mahla, Swarnim Science College

**Co-investigator:** Dhara Goti, Darshika Savalia, Narmatta Datta ,Priyanka Shrimali, Mitesh Prajapati

1. I/We agree to abide by the terms and conditions of the grant.

2. I/We did not submit this or a similar project proposal elsewhere for financial support. I/we have explored and ensured that equipment and basic facilities will actually be available as and when required for the purpose of the project.

3. I/We shall not request financial support under this project for procedure for procurement of these items.

4. I/We undertake that spare time on permanent equipment and will be made available to other Researchers/users.

5. I/We undertake to submit progress reports, Statement of Expenditure (SE)/ accounts, Utilisation Certificates (UC) etc covering upto 31<sup>st</sup> March every year till completion of the project as prescribed by DST.

6. I/We have enclosed the following materials:

- a. Endorsement from the Head of the Institution (on letter head)
- b. Certificate from Investigator (s)
- c. Completed Proposal one hard copy

Date:

  
Signature of PI/PI's

Place: *SSIU, Gandhinagar*

## ENDORSEMENT FROM THE HEAD OF THE INSTITUTION

**PROJECT TITLE:** Copper Biosorption Utilizing Dehydrated Aspergillus Biomass

**Name(s) of the PI's and Co-PI's:**

**Principal Investigator:** Dr. Priti Mahla, Assistant Professor, Swarnim Science College

**Co-investigator:** Dhara Goti, Darshika Savalia, Narmatta Datta ,Priyanka Shrimali, Mitesh Prajapati

It is certified that the Institute welcomes participation of above as the Principal Investigator(s) and/or Co-PI(s) for the project. The research proposal falls in line with the normal research activities of the Institution.

27. Certified that the equipment other basic facilities and such other administrative and accounting facilities as per terms and conditions of the grant will be extended to the investigator throughout the duration of project. In case of PI/Co-PI leaving the institution or proceeding on a long leave during the project period, a prior permission will be sought from DST before relieving/sanctioning.
28. Institute assumes the financial and other management responsibilities of the project including timely submission of Progress Reports, UC's and SE's and facilitate in conducting external audit, if required.

Name of the Head of Institution

Signature with stamp

Date:

Place: SSIU, Gandhinagar



SWARNIM  
STARTUP & INNOVATION  
UNIVERSITY

## Declaration by Principal Investigator

I Dr. Priti Mahla hereby declare that the **major research Project Copper Biosorption Utilizing Dehydrated Aspergillus Biomass** is supported by Spechrom Solutions is submitted to Swarnim Startup and Innovation University. This is my original work and my indebtedness to other work publications, references, if any, have been duly acknowledged. If I am found guilty of copying from any other report or published information and showing as my original work, I understand that I shall be liable and punishable by the university.

Signature:

Name of PI: Dr. Priti Mahla

Designation: Assistant Professor

Name of Department: Microbiology

Name of Institute with code: Swarnim Science College

Stamp of Institute

Verified By

Principal:  
Signature





# **Copper biosorption utilizing dehydrated *Aspergillus* biomass**

**Research Project Report Submitted to**

**Spechrom Solution Pvt Lmt.**



**Submitted by:**

**Principal Investigator:** Dr. Priti Mahla, Head, Assistant Professor, Swarnim  
Science College

**Co-investigator:** Dhara Goti, Darshika Savalia, Narmatta Datta, Priyanka Shrimali,  
Mitesh Prajapati

## INDEX

Sr. No	Content	Page No
1	Introduction	
2.	Literature Review	
3.	Rationale	
4.	Aim and Objectives	
5.	Methodology	
6.	Result and Discussion	
7	Conclusion	
8.	References	

## 1. Introduction:

The issue of environmental contamination has become a primary concern for all the nations on a global scale, as it affects and causes suffering in both developed and developing countries. The pollution of water resources is a significant concern among all environmental pollutants. The presence of heavy metals in sewage water is a result of the increased use of these substances in industrial processes. (Amorim, et al., 2018) (Andreazza, et al., 2010) Heavy metals are the most hazardous type of pollution since they don't biodegrade and remain for long in the environment. (Lebron, Moreira, Santos, & Jacob, 2018) (Gupta, Rastogi, Saini, & Jain, 2006) (de Carvalho, et al., 2003) The presence and accumulation of metallic ions in industrial effluent are a cause for major concern due to their deleterious effects on living organisms (Areco & dos Santos Afonso, 2010) (Baltazar, et al., 2018).

Copper is one of the most widespread environmental contaminants, which can be toxic to living organisms. Copper is released into the environment during the mining of copper and other metals, as well as from enterprises that produce or utilize copper metal. Additional entry points include wood production, natural sources, and home waste water and fossil fuel ingestion. (Mathew & Krishnamurthy, 2018)

Toxic metals such as copper is removed from waste waters using a variety of physico-chemical techniques, like reverse osmosis, Ion exchange, Ultrafiltration, Precipitation, Adsorption, Absorption, Activated carbon absorption, Membrane filtration, Bioleaching, Membrane bioreactor, Biosorption. Among them Biosorption inexpensive biomaterials to sequester environment pollutants from aqueous solutions by a wide range of Physico-chemical mechanism, including; ion exchange, chelation, complexation, physical adsorption and surface microprecipitation. Biosorption is a quick, mainly reversible, passive absorption mechanism that doesn't depend on the vitality of the cell. (Nazari, Cox, & Waters, 2014). As metal ions can accumulate and cause cancer in living things, there has been a lot of research done on the bio-adsorption technique for discharging metal ions from industrial wastewater. (Baltazar, et al., 2018) (Barquilha, Cossich, Tavares, & Silva, Biosorption of nickel (II) and copper (II) ions by *Sargassum* sp. in nature and alginate extraction products, 2019).



Fungi are another type of microorganisms that have been efficiently exploited in the preparation of biosorbent for metal ions removal from polluted soil. Fungi are easy to grow, provide significant quantities of biomass and at the same time can be altered genetically and morphologically (Kapoor et al., 1995). Different types of fungi can break down different levels of pollutants because they can release extracellular enzymes that help with the assimilation of complex carbohydrates for earlier hydrolysis. The benefit lies in the easy separation of fungal biomass by filtration, facilitated by its filamentous structure. Fungal biomass may be utilized by biosorptive processes as it often exhibits marked tolerance towards metals and other factors such as low pH (Gadd, 1990; Zafar et al., 2007; Kapoor and Viraraghavan, 1997). Fungal cell walls and their components have a major role in biosorption. Many fungal species such as *Rhizopus arrhizus*, *Penicillium spinulum* and *Aspergillus niger* have been extensively studied for heavy metal biosorption and the mechanism seems to be species dependent (Zhou and Kiff, 1991; Hafez et al., 1997). (Nur Liyana Iskandar1 Tolerance and biosorption of copper (Cu) and lead (Pb) by filamentous fungi isolated from a freshwater ecosystem).

In this study fungi were isolated from different samples such as soil, leaves and decayed food collected from Ahmedabad and Aqueous copper waste collected from Vatva GIDC. The maximum biosorption capacity based on dry weight was determined by varying the concentrations of Cu ions in the aqueous solution.

## 2. Literature Review

Treatability studies have indicated that wastes often contain toxic and non-biodegradable matter (such as metals), which generally limits the implementation of a conventional biological treatment process. The mining, mineral processing and metallurgical industries are known to generate annually billions of tons of wastes. Some of the metal and mineral constituents of these wastes have potential economic value and their recovery can augment our primary mineral and metal resource supply base. The important of pollution prevention by clean technologies instead of remediation has also been recognized.

The U.S. Environmental Protection Agency (EPA) is evaluating technologies for their

ability to recover metals from sludges and wastewater. Dilute aqueous solutions, generated or used by industry, can contain a variety of different metal ions. Various processes are suitable for reclamation of toxic metals and among them, attention is paid here to biosorption. The ability of microorganisms to remove metal ions from solution is a well-known phenomenon.

Industrial applications of biosorption often make use of dead biomass, which does not require nutrients and can be exposed to environments of high toxicity. Experimental laboratory batch experiments are described for fungi and for activated sludge, as the metal biosorbents, providing insight into copper biosorption. Non-living biomass showed greater binding capacities for copper (a priority pollutant) than living biomass. Engineering considerations are central in decisions concerning the commercial future of biosorption and a practical solution is needed for certain problems, such as the efficient separation of metal-loaded biomass.

Pollution of surface and ground water caused by human and industrial activities has been recorded as a major problem in the global context ( Satya et al.,2011) water pollution considered as the leading universal cause of 80% of diseases ( OECD, 2006). The presence of inorganic pollutants such as metal ions in the ecosystem causes a major environmental problem. Toxic metal compounds coming to the earth's surface, it contaminate earth's water as well as underground water in trace amounts by leaking from the soil after rain and snow( Kilic et al.).

Metal ions are reported as priority pollutants due to their mobility, in natural water ecosystems and due to their toxicity (Demirbas A.,2008). The problem associated with metal ion pollution is that they are not biodegradable and are highly persistent in the environment. Thus, they can be accumulated in living tissues, causing various disease and disorders ( Wan Ngah WS, 2008). Heavy metal toxicity can result in damage or reduced mental and central nervous function, lower energy levels and damage to blood composition, lungs, kidneys, liver and other vital organs ( Ahmaruzzaman M.,2011).

The term bioremediation has been introduced to describe the process of using biological agents to remove toxic waste from environment. Bioremediation is most



effective management tool to manage the polluted environment, and recover contaminated soil and water. Bioremediation has been used at a number of sites worldwide with varying degrees of success.(Kumar. A 2011).Intensification of agriculture and manufacturing industries has resulted in increased release of a wide waste has been led to scarcity of clean water and disturbances of hazardous waste has been led to scarcity of clean water and distribution of soil thus limiting crop production (Kumaluden et al.,2003).

This technology relies on promoting the growth of specific microflora or microbial consortia that are indigenous to the contaminated sites that are able to perform desired activities (Agarwal,1998). Mycoremediation is a form of bioremediation in which fungi-based technology is used to decontaminate the environment. Phrase coined by Paul Stamets.Fungi have been proven to be a very cheap, effective and environmentally sound way for helping to remove a wide array of toxins (heavy metals) from damage environment like soil.

Biosorption is a physico-chemical process, simply defined as the removal of substances from by biological material. Biosorption uses inexpensive biomaterial to sequester environment pollutants from aqueous solutions by a wide range of physico-chemical mechanism, including; ion exchange, chelation, complexation, physical adsorption and surface microprecipitation. The biomaterials used in this process are termed biosorbent (flores-Garnica JG et al.,2013).

Fungi represent another group of microorganisms that have been efficiently used in the preparation of biosorbent for metal ions removal from contaminated soil. Fungi are easy to grow, produce high yields of biomass and at the same time can be manipulated genetically and morphologically (Kapoor et al..1995). Therefore, the aim of this review is to discuss about the heavy metal removal from contaminated sources and aqueous solutions by fungi of different groups. The capability of many type of fungi to produce extracellular enzymes for assimilation of complex carbohydrates for former hydrolysis makes capable the degradation of various degrees of pollutants. Benefit is easy separation of fungal biomass by filtration because of its filamentous structure.



The bio-sorptive capacity of dead fungal biomass has been studied by several authors during the last few years. A.Kapoor & T. Viraraghavan\* reported a high biosorption capacity of *Aspergillus* sp. Javaid et al.explored the adsorption potential of *Pleurotus ostreatus* to remove copper from water and achieved maximum biosorption capacities of 8.06 mg g<sup>-1</sup> .

### 3. Rationale:

The rationale behind selecting *Aspergillus* biomass lies in its filamentous structure, which provides a large surface area for biosorption and allows for easy separation from the treated water. The study's focus on dehydrated biomass is particularly important, as it explores the potential for long-term storage and reusability of the biosorbent, thus enhancing the practical applicability of this method in real-world scenarios.

The use of FTIR analysis to identify the functional groups involved in the biosorption process provides a deeper understanding of the mechanisms at play, which can further inform the development of more efficient biosorbents in the future. By demonstrating the effectiveness of *Aspergillus* biomass in copper biosorption, this study contributes to the growing body of research on sustainable and cost-effective methods for heavy metal removal, with potential implications for improving water quality and protecting public health.

### 4. Aim and Objectives

The study was aimed to provide a practical view of the process involved in biosorption dealing with the copper contaminated water using *Aspergillus*.

1. Collection of aqueous waste
2. Isolation of Fungus
3. Screening of fungi for copper sorption
4. Optimization of system using different parameters

### 5. Methodology:

### **5.1 Sample collection:**

Aqueous Copper sample was collected from Industry named Selex, G I D C, Phase 2, Vatva, Ahmedabad, Gujarat, 382445.

### **5.2 Isolation and Characterization**

Sterile Potato Dextrose Agar plates were used for the isolation of various fungi from sources like soil, decay food, leaf. The Streak plate method was performed and plates were incubated for 6/7 days at room temperature. Lactophenol blue was used for mounting of fungus.

### **5.3 Biomass production**

Biomass production was done in 100ml sterile PDA broth by keeping the flasks under shaking conditions at 150 rpm or a week.

### **5.4 Process of Biosorption**

Batch experiment was done. Biosorption study were performed in 250 ml Erlenmeyer flask by addition of adsorbent to 50 ml copper solution. The pH was adjusted with 1 N HCL and 1 N NaOH before addition of biosorbent(biomass). In all the experiments, the flask was shaken at 150 rpm on a rotary shaker. Contact time was 2 hours. At different interval of time, the samples were withdrawn and filtered. The residual metal ion in the filtrate was extracted using solvent extraction method and was determined spectrophotometrically. Control system was prepared without adding the biomass.

### **5.5 Optimization of Process Parameter**

The optimization of various parameters was done like the effect of biomass concentration, pH, agitation. For agitation, amount of bio-sorbent and pH were constant (pH 4.5, biosorbent concentration 80 mg). The effect of bio-sorbent was determined using adsorption of Cu (II) 10-80 mg/100 ml and other experimental conditions were fixed (copper solution concentration 100 ppm / 100 mg pH 4.5). The effect of pH on the copper adsorption was studied at different pH values (2.5-5.5) at constant copper concentration (100 ppm/100 ml), amount of bio-sorbent (80 mg/100 ml).

### **5.6 Calculation of The efficiency of adsorbed copper was calculated as follows:**

Efficiency of adsorption(%) =  $\frac{E_0 - E}{E_0} \times 100$  [Where,  $E_0$  = Initial metal ion concentration (O.D.)  $E$  = Final metal ion concentration (O.D.)]

### **5.7 Solvent extraction**

Working solution of cupric sulphate pentahydrate was prepared from the stock solution by diluting it to get desired concentration and then biomass was added to it. 1.0 ml from the experimental system was added into beaker. 10.0 ml of distilled water and 5.0 ml of 25% aqueous citric acid solution was added to it. It was rendered slightly alkaline with ammonia solution, alternatively adjusted to pH 8.5. Then, it was added with 15 ml of 4% EDTA solution and was transferred to a separatory funnel. 0.2% aqueous SDDC (sodium diethyldithiocarbamate) solution was added and then shaken well for 45 second or given 90 strokes, a yellow brown color developed in the solution. Further 10.0 ml of n-Butyl acetate was added into the funnel and shaken for 45 second again or given 90 strokes. The phases were allowed to get separated, the lower aqueous layer was removed. 10 ml of 10% sulfuric acid was allowed to get separated and the lower aqueous layer was discarded. The optical density was determined at 560 nm spectrophotometrically.

### **5.8 FTIR Analysis:**

Functional groups of the adsorbent material were confirmed by performing FTIR.

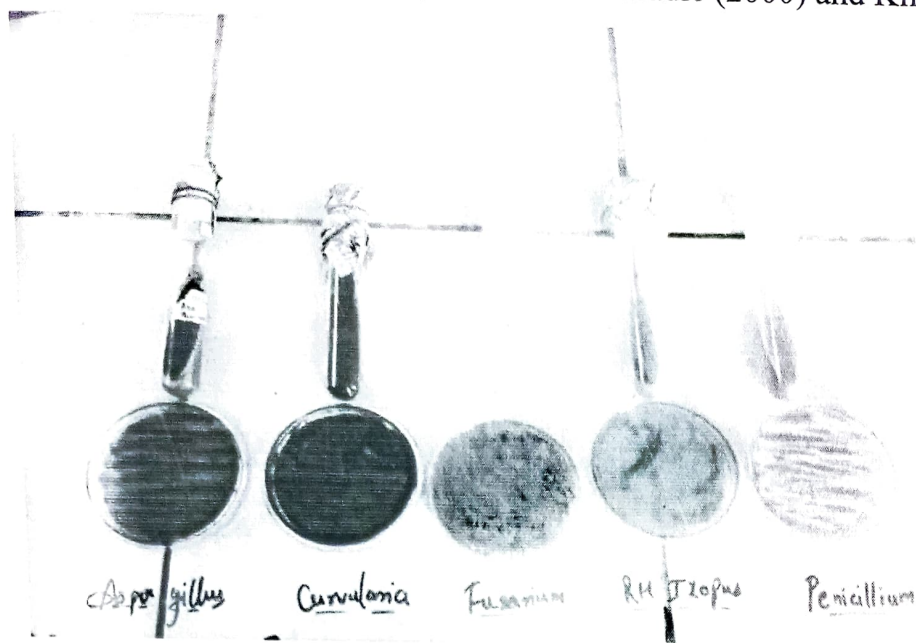
## **6. Result and Discussion**

### **6.1 Fungal Isolation**

Total five isolates of fungi were obtained from soil, decayed food, leaves, etc., and they were named *Aspergillus*, *Curvularia*, *Fusarium*, *Rhizopus*, and *Penicillium*. *Aspergillus* fungal biomass was used for further work. The cultural characters of the *Aspergillus* showed dry, powdery growth with shades like black, green, brown, and yellow, which is consistent with previous reports on its growth pattern (Klich, 2002). The morphological characteristics of *Aspergillus* included septate mycelia, with conidiophores that were long, unbranched, and ending in vesicles, on which sterigmata were radially arranged in chains, as



described by Moss and Cheadle (2000) and Klich (2002).



**Figure:** different five types of fungal isolates in PDA plate and pure culture in PDA slant.



**Figure:** microscopic observation of *Aspergillus* sp.

## 6.2 EFFECT OF BIOSORBENT (BIOMASS)

The removal of Cu (II) ions from aqueous solution was significantly dependent on the amount of biosorbent. The result shown in the graph indicates that the sorption increased up to a certain limit and then decreased. The increased biosorption with the biosorbent concentration can be attributed to greater surface area and the availability of more biosorption sites (Volesky, 2007). At biosorbent concentrations greater than 40 mg, the surface Cu (II) concentration and the solution Cu (II) concentration came to equilibrium with each other. However, the biosorption capacity decreased with an increase in biosorbent concentration. This may be attributed to overlapping or aggregation of biosorbent sites, resulting in a decrease in the total biosorbent surface area available to Cu (II) ions (Aksu & Tezer, 2005).

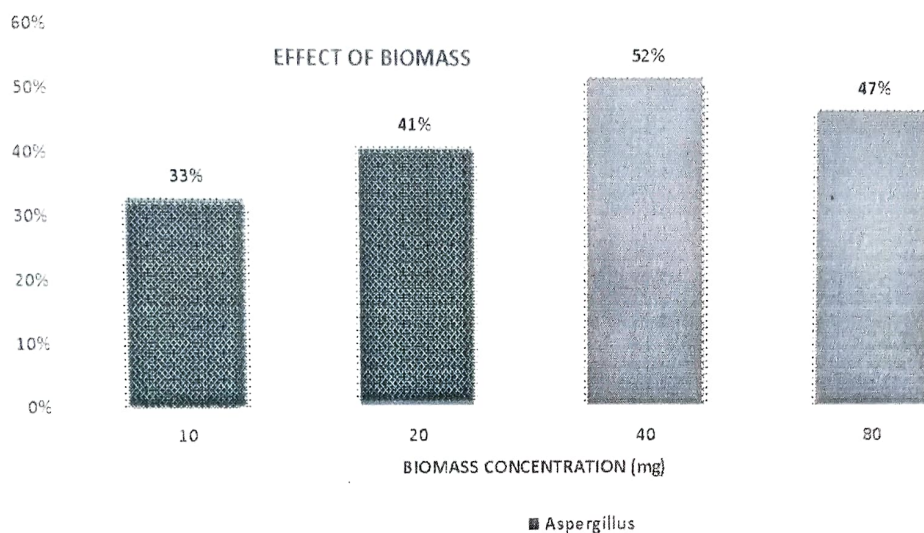
Biomass Concentration (mg)	Copper Removal Efficiency (%)
10	33%
20	41%
40	52%
80	47%

The efficiency of copper ion removal was observed to vary significantly with changes in the biomass concentration of dehydrated *Aspergillus* used in the biosorption process. The data showed a positive correlation between biomass concentration and biosorption efficiency up to a certain point, after which a slight decline was noted. At a biomass concentration of 10 mg, the copper removal efficiency was recorded at 33%. Increasing the biomass to 20 mg led to a noticeable increase in biosorption, achieving 41% efficiency. The highest removal efficiency was obtained at 40 mg, where copper biosorption peaked at 52%. This enhancement in metal uptake can be attributed to the increased availability of active binding sites on the fungal cell wall as more biosorbent was introduced into the system (Volesky, 2007). However, further increasing the biomass concentration to 80 mg resulted in a reduction of biosorption efficiency to 47%. This slight decrease may be due to overlapping or aggregation of biosorbent particles at higher concentrations, which causes a reduction in the effective surface area available for copper ion binding (Aksu & Tezer, 2005). Additionally, excess biomass in a fixed volume of solution may lead to saturation of binding sites and hinder optimal mass transfer between the solute and biosorbent (Anand et al., 2020).

These findings suggest that there is an optimum biomass concentration (around 40 mg) for achieving maximum biosorption efficiency. Beyond this point, additional biomass does not contribute proportionally to metal ion removal and may even interfere with the process. This trend is consistent with biosorption behaviors reported in the literature, where optimal biosorbent dosage balances the availability of functional groups and mass transfer dynamics (Meena et al., 2008). The functional groups (e.g., hydroxyl, carboxyl, and amino groups) on the surface of *Aspergillus* biomass are primarily responsible for metal ion uptake through ion exchange, chelation, or physical adsorption mechanisms (Pino et al., 2006).

The experimental results demonstrate that biomass concentration is a critical factor influencing the efficiency of copper biosorption. 40 mg of biomass was found to be the most effective dosage under the studied conditions, highlighting the need to optimize biosorbent amounts for practical applications in wastewater treatment.

Graph: Effect of concentration of biomass at pH 4.5



At 40 mg of biomass concentration, maximum sorption (52%) was achieved. (A.verma et 2013). Increase in biosorbent concentration is linked with the decrease in biosorbent capacities. This may be due to aggregation sites on fungi, causing a decrease in total adsorbent surface area available to Cu (II) (Din and Mirza, 2013).

### EFFECT OF AGITATION

Results of the experiments showed that biosorption is increased in shaking condition compared to static condition.

Mode of Agitation	Biosorption (%)
Static	20%
Shaking	52%

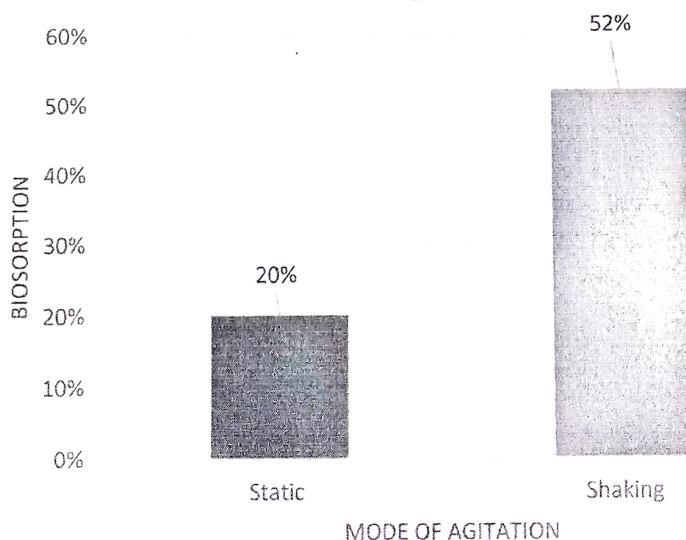


The mode of agitation significantly influenced the biosorption efficiency of copper ions using dehydrated *Aspergillus* biomass. Two different conditions were analyzed: static (no shaking) and shaking (agitated). Under static conditions, the copper biosorption efficiency was relatively low, recorded at 20%. In contrast, when the system was subjected to shaking, the biosorption efficiency increased markedly to 52%. This substantial improvement can be attributed to enhanced contact between the metal ions and the biosorbent surface (Aksu & Tezer, 2005).

Agitation facilitates better dispersion of the biomass in the solution, thereby reducing boundary layer resistance and increasing the rate of mass transfer of copper ions from the bulk solution to the biosorbent surface (Volesky, 2007). This results in improved accessibility of active binding sites on the fungal biomass. Shaking ensures that the solution remains homogeneous, preventing localized saturation of metal ions around biomass particles and promoting more uniform uptake (Pino et al., 2006). On the other hand, in static conditions, the lack of movement limits the interaction between the biosorbent and the metal ions, resulting in lower biosorption efficiency. The reduced diffusion of copper ions toward the biosorbent surface leads to a slower uptake rate and fewer ions being adsorbed within the given contact time (Anand et al., 2020).

The experimental results clearly demonstrate that agitation (shaking) enhances biosorption efficiency significantly, making it a vital operational parameter to consider for optimizing the removal of heavy metals such as copper from aqueous solutions using fungal biomass (Meena et al., 2008). These findings underscore the importance of agitation in biosorption processes. Proper agitation not only improves the kinetics of biosorption but also ensures that the maximum number of binding sites are utilized effectively. This is particularly relevant for large-scale applications where achieving high efficiency in a shorter time is critical.

### Effect of Agitation on Biosorption





Graph: Effect of Agitation at pH 4.5; where, efficiency of sorption is 20% in static condition and 52% in shaking condition

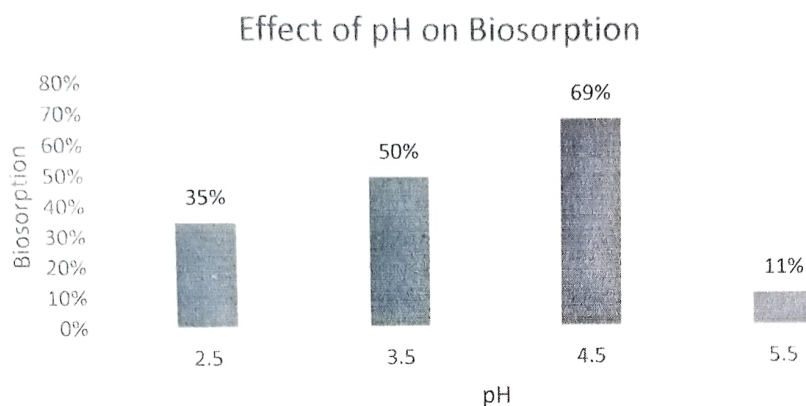
### 6.3 Effect Of pH

The result of the experiment showed that biosorption was low in a strong acidic medium, and adsorption capacity increased with an increase in pH values, up to a certain pH level. Biosorption was enhanced between pH 2 and 5. The results are shown in the graph. At the optimal pH of 4.5, 59% of Cu(II) ions were removed. At pH values above 5, copper ions in the solution precipitated, as shown in the graph. At low pH, the H<sup>+</sup> ions competed with metal cations (Cu(II)) for the exchange sites on the biosorbent, thereby partially hindering metal uptake under conditions of extreme acidity (Aksu & Tezer, 2005). The competition between H<sup>+</sup> ions and Cu(II) ions for the binding sites on the biosorbent is a well-known phenomenon that reduces biosorption efficiency at lower pH levels (Volesky, 2007)

pH	Biosorption (%)
2.5	35%
3.5	50%
4.5	69%
5.5	11%

The result of the experiment showed that biosorption was low in a strong acidic medium, and adsorption capacity increased with an increase in pH values, up to a certain pH level. Biosorption was enhanced between pH 2 and 5. The results are shown in the graph. At the optimal pH of 4.5, 59% of Cu(II) ions were removed. At pH values above 5, copper ions in the solution precipitated, as shown in the graph. At low pH, the H<sup>+</sup> ions competed with metal cations (Cu(II)) for the exchange sites on the biosorbent, thereby partially hindering metal uptake under conditions of extreme acidity (Aksu & Tezer, 2005). The competition between H<sup>+</sup> ions and Cu(II) ions for

the binding sites on the biosorbent is a well-known phenomenon that reduces biosorption efficiency at lower pH levels (Volesky, 2007).



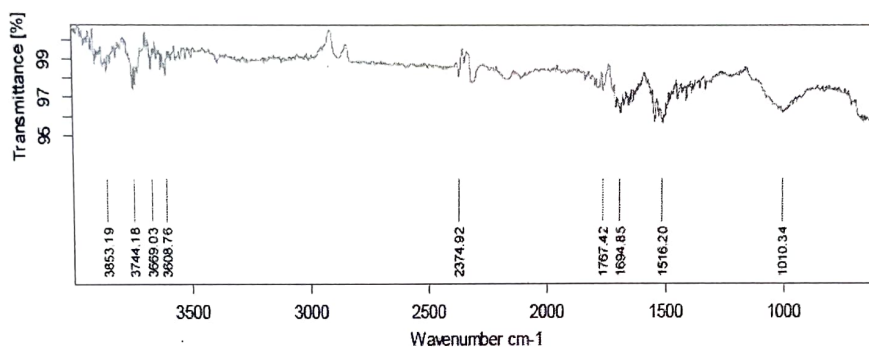
Graph: Effect of pH at Cu(II) ion concentration of 50 ppm and biosorbent concentration of 80 mg.

#### 6.4 FTIR Analysis

In order to determine the characteristic functional groups responsible for biosorption of copper ions, FTIR spectroscopy was applied. The bonding mechanism between copper and biomass can be determined by interpreting the infrared absorption spectrum (Volesky, 2007). These results revealed that a chemical interaction between the copper ions and the hydroxyl groups occurred on the biomass surface (Aksu & Tezer, 2005). From the analysis of the IR spectra, it can be seen that the mechanism of copper uptake by fungal biomass involves interactions between copper ions and hydroxyl, amino, carboxyl, and carbonyl groups from the fungal biomass surface, depending on the types of fungal strain (Pino et al., 2006; Meena et al., 2008).

Wavenumber (cm <sup>-1</sup> )	Probable Functional Group / Vibration
385	O-H stretching (alcohols/phenols, hydrogen bonding)
3744.18	Free O-H stretch (possibly alcohol or water)
3696.03	O-H/N-H stretching (weak hydrogen bonding)

3680.76	O-H stretching vibrations
2374.92	C≡N stretching (nitriles), CO <sub>2</sub> overtones (less likely)
1767.42	C=O stretching (esters, anhydrides)
1694.85	C=O stretching (carboxylic acids, amides)
1516.20	N-O asymmetric stretch (nitro compounds) or aromatic C=C stretch
1010.34	C-O stretching or polysaccharide region (alcohols, ethers)



D:\FTIR\Aspergillus.0		Aspergillus	Instrument type and / or accessory			
Hit No	Hit Quality	Compound Name	Entry No.	Lib. Index	Molecular formula	Molecular weight CAS number
1	233	Amides - Tertiary	31	1		
2	112	Ethers - Aromatic	19	1		
3	34	Nitriles - Aromatic	39	1		
4	23	Alcohols - Isomolstr	15	1		
5	13	Amides - Secondary	30	1		
6	9	Nitriles - Aliphatic	38	1		
7	8	Amides - Primary	29	1		
8	7	Alkyls-Disub	6	1		
9	7	Nitro Compounds - Nitrites	45	1		
10	3	Isonitriles Aromatic	41	1		

### BUDGET UTILIZED:

Category	Item Description				Total (INR)
		1 Year	2 Year	3 Year	
<b>Chemicals &amp; Reagents</b>	Copper sulfate, acids, buffers, and other reagents	9000		6000	15,000
<b>Biomass Preparation</b>	Aspergillus culture, growth media,	50000		10000	60,000



	dehydration setup				
<b>Laboratory Equipment</b>		15000			15,000
<b>Instrumentation</b>	AAS/ICP for copper analysis	55000	105000		160,000
<b>Microbial Culturing</b>	Incubators, autoclave usage, agar plates	50000			50,000
<b>Utilities &amp; Consumables</b>	Distilled water, electricity, gloves, disposable items	14000		10000	24,000
<b>Data Analysis Software</b>	MATLAB/SPSS/Excel for statistical analysis	15000			15,000
<b>Contingency Fund</b>	Unforeseen expenses	10000		10000	20,000
<b>Total (INR)</b>		<b>309000</b>		<b>50000</b>	<b>359000</b>

## 7. Conclusion:

Hence, from the study it was concluded that the fungi are more helpful for sorption of copper from aqueous copper solution as compared to all other biomasses. Bio-sorption is rapid and achieved at small time interval within an hour of experiment. Many problems arise from copper contaminated water. Therefore, an economically viable and easy method for bio-sorption of copper from aqueous solution is highly desirable.

## 8. References

1. Kapoor, A., & Viraraghavan, T. (1998). Biosorption of heavy metals on aspergillus niger. Effect of pretreatment. *Bioresources Technology*, 63(2), 109-113. Doi: 10.1016/s0960-8524(97)00118-1
2. Innocent OBOH\*, Emmanuel ALUYOR, and Thomas AUDU, Biosorption of Heavy Metal Ions from Aqueous Solutions Using a Biomaterial, Issue 14, January-June 2009 p. 58-65,
3. N. Ahalya, T.V. Ramachandra\*<sup>1</sup> and RD. Kanamadi<sup>2</sup>, Biosorption of Heavy Metals, Vol.7 (4) Dec. (2003) Res. J. Chem. Environ
4. Al-Homaidan, A., Al-Houri, H., Al-Hazzani, A., Elgaaly, G., & Moubayed, N. (2014). Biosorption of copper ions from aqueous solutions by *Spirulina platensis* biomass. *Arabian Journal of Chemistry*, 7(1), 57-62.
5. Al-Rub, F., El-Naas, M., Ashour, I., & Al-Marzouqi, M. (2006). Biosorption of copper on *Chlorella vulgaris* from single, binary and ternary metal aqueous solutions. *Process Biochemistry*, 41(2), 457-464.
6. Amorim, S., Ruas, F., Barboza, N., de Oliveira Neves, V., Leão, V., & Guerra-Sá, R. (2018). Manganese (Mn<sup>2+</sup>) tolerance and biosorption by *Meyerozyma guilliermondii* and *Meyerozyma caribbica* strains. *Journal of Environmental Chemical Engineering*, 6(4), 4538-4545.
7. Andreazza, R., Pieniz, S., Wolf, L., Lee, M.-K., Camargo, F., & Okeke, B. (2010). Characterization of copper bioreduction and biosorption by a highly copper resistant bacterium isolated from copper-contaminated vineyard soil. *Science of The Total Environment*, 408(7), 1501-1507.
8. Areco, M., & dos Santos Afonso, M. (2010). Copper, zinc, cadmium and lead biosorption by *Gymnogongrus torulosus*. Thermodynamics and kinetics studies. *Colloids and Surfaces B: Biointerfaces*, 81(2), 620-628.
9. Baltazar, M., Gracioso, L., Avanzi, I., Karolski, B., Tenório, J., do Nascimento, C., & Perpetuo, E. (2018). Copper biosorption by *Rhodococcus erythropolis* isolated from the Sossego Mine – PA – Brazil. *Journal of Materials Research and Technology*.
10. Bano, A., Hussain, J., Akbar, A., Mehmood, K., Anwar, M., Hasni, M., . . . Ali, I.

- (2018). Biosorption of heavy metals by obligate halophilic fungi. *Chemosphere*, 199, 218-222.
11. Barquilha, C., Cossich, E., Tavares, C., & Silva, E. (2017). Biosorption of nickel(II) and copper(II) ions in batch and fixed-bed columns by free and immobilized marine algae *Sargassum* sp. *Journal of Cleaner Production*, 150, 58-64.
  12. Blázquez, G., Martín-Lara, M., Dionisio-Ruiz, E., Tenorio, G., & Calero, M. (2011). Evaluation and comparison of the biosorption process of copper ions onto olive stone and pine bark. *Journal of Industrial and Engineering Chemistry*, 17(5), 824-833.
  13. Blázquez, G., Martín-Lara, M., Dionisio-Ruiz, E., Tenorio, G., & Calero, M. (2012). Copper biosorption by pine cone shell and thermal decomposition study of the exhausted biosorbent. *Journal of Industrial and Engineering Chemistry*, 18(5), 1741-1750.
  14. Calero, M., Iáñez-Rodríguez, I., Pérez, A., Martín-Lara, M., & Blázquez, G. (2018). Neural fuzzy modelization of copper removal from water by biosorption in fixed-bed columns using olive stone and pinion shell. *Bioresource Technology*, 252, 100-109.
  15. da Rosa, A., Carissimi, E., Dotto, G., Sander, H., & Feris, L. (2018). Biosorption of rhodamine B dye from dyeing stones effluents using the green microalgae *Chlorella pyrenoidosa*. *Journal of Cleaner Production*, 198, 1302-1310.
  16. Dang, V., Doan, H., Dang-Vu, T., & Lohi, A. (2009). Equilibrium and kinetics of biosorption of cadmium(II) and copper(II) ions by wheat straw. *Bioresource Technology*, 100(1), 211-219.
  17. do Nascimento, J., de Oliveira, J., Rizzo, A., & Leite, S. (2019). Biosorption Cu (II) by the yeast *Saccharomyces cerevisiae*. *Biotechnology Reports*, 21, e00315.
  18. Dong, Y., Lin, H., Xu, X., Zhang, Y., Gao, Y., & Zhou, S. (2013). Comparative study on the bioleaching, biosorption and passivation of copper sulfide minerals. *International Biodeterioration & Biodegradation*, 84, 29-34.
  19. Ekmekyapar, F., Aslan, A., Bayhan, Y., & Cakici, A. (2006). Biosorption of copper(II) by nonliving lichen biomass of *Cladonia rangiformis* hoffm. *Journal of Hazardous Materials*, 137(1), 293-298.
  20. Fadel, M., Hassanein, N., Elshafei, M., Mostafa, A., Ahmed, M., & Khater, H. (2017).



- Biosorption of manganese from groundwater by biomass of *Saccharomyces cerevisiae*. *HBRC Journal*, 13(1), 106-113.
21. Figueroa-Torres, G., Certucha-Barragán, M., Acedo-Félix, E., Monge-Amaya, O., Almendariz- Tapia, F., & Gasca-Estefanía, L. (2016). Kinetic studies of heavy metals biosorption by acidogenic biomass immobilized in clinoptilolite. *Journal of the Taiwan Institute of Chemical Engineers*, 61, 241-246.
  22. Flouty, R., & Estephane, G. (2012). Bioaccumulation and biosorption of copper and lead by a unicellular algae *Chlamydomonas reinhardtii* in single and binary metal systems: A comparative study. *Journal of Environmental Management*, 111, 106-114.
  23. Hsueh, Y.-H., Lin, K.-S., Wang, Y.-T., & Chiang, C.-L. (2017). Copper, nickel, and zinc cations biosorption properties of Gram-positive and Gram-negative MerP mercury-resistance proteins. *Journal of the Taiwan Institute of Chemical Engineers*, 80, 168-175.
  24. Hu, H., Zhang, J., Lu, K., & Tian, Y. (2015). Characterization of *Acidosasa edulis* shoot shell and its biosorption of copper ions from aqueous solution. *Journal of Environmental Chemical Engineering*, 3(1), 357-364.
  25. Izquierdo, M., Gabaldón, C., & Marzal, P. (2014). Modeling of the effect of EDTA on copper(II) biosorption onto *Posidonia oceanica* waste in batch and fixed-bed systems. *Journal of the Taiwan Institute of Chemical Engineers*, 45(2), 665-673.
  26. Izquierdo, M., Gabaldón, C., Marzal, P., & Álvarez-Hornos, F. (2010). Modeling of copper fixed-bed biosorption from wastewater by *Posidonia oceanica*. *Bioresource Technology*, 101(2), 510-517.
  27. Jianlong, W. (2002). Biosorption of copper(II) by chemically modified biomass of *Saccharomyces cerevisiae*. *Process Biochemistry*, 37(8), 847-850.
  28. Kaduková, J., & Virčíková, E. (2005). Comparison of differences between copper bioaccumulation and biosorption. *Environment International*, 31(2), 227-232.
  29. Kariuki, Z., Kiptoo, J., & Onyancha, D. (2017). Biosorption studies of lead and copper using rogers mushroom biomass '*Lepiota hystrix*'. *South African Journal of Chemical Engineering*, 23, 62-70.
  30. Keshtkar, A., Moosavian, M., Sohbatzadeh, H., & Mofras, M. (2019). La(III) and

- Ce(III) biosorption on sulfur functionalized marine brown algae *Cystoseira indica* by xanthation method: Response surface methodology, isotherm and kinetic study. *Groundwater for Sustainable Development*, 8, 144-155.
31. Khormaei, M., Nasernejad, B., Edrisi, M., & Eslamzadeh, T. (2007). Copper biosorption from aqueous solutions by sour orange residue. *Journal of Hazardous Materials*, 149(2), 269-274.
  32. Letnik, I., Avrahami, R., Port, R., Greiner, A., Zussman, E., Rokem, J., & Greenblatt, C. (2017). Biosorption of copper from aqueous environments by *Micrococcus luteus* in cell suspension and when encapsulated. *International Biodeterioration & Biodegradation*, 116, 64-72.
  33. Liu, X., Chen, Z.-Q., Han, B., Su, C.-L., Han, Q., & Chen, W.-Z. (2018). Biosorption of copper ions from aqueous solution using rape straw powders: Optimization, equilibrium and kinetic studies. *Ecotoxicology and Environmental Safety*, 150, 251-259.
  34. Londono-Zuluaga, C., Jameel, H., Gonzalez, R., & Lucia, L. (2019). Crustacean shell-based biosorption water remediation platforms: Status and perspectives. *Journal of Environmental Management*, 231, 757-762.
  35. Long, J., Gao, X., Su, M., Li, H., Chen, D., & Zhou, S. (2018). Performance and mechanism of biosorption of nickel(II) from aqueous solution by non-living *Streptomyces roseorubens* SY. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 548, 125-133.
  36. Lopez-Fernandez, M., Moll, H., & Merroun, M. (2019). Reversible pH-dependent curium(III) biosorption by the bentonite yeast isolate *Rhodotorula mucilaginosa* BII-R8. *Journal of Hazardous Materials*, 370, 156-163.
  37. Markou, G., Mitrogiannis, D., Çelekli, A., Bozkurt, H., Georgakakis, D., & Chrysikopoulos, C. (2015). Biosorption of  $\text{Cu}^{2+}$  and  $\text{Ni}^{2+}$  by *Arthrospira platensis* with different biochemical compositions. *Chemical Engineering Journal*, 259, 806-813.
  38. Martín-Lara, M., Blázquez, G., Calero, M., Almendros, A., & Ronda, A. (2016). Binary biosorption of copper and lead onto pine cone shell in batch reactors and in

fixed bed columns. *International Journal of Mineral Processing*, 148, 72-82.

39. Mata, Y., Blázquez, M., Ballester, A., González, F., & Muñoz, J. (2009). Biosorption of cadmium, lead and copper with calcium alginate xerogels and immobilized *Fucus vesiculosus*. *Journal of Hazardous Materials*, 163(2), 555-562.
40. Mata, Y., Blázquez, M., Ballester, A., González, F., & Muñoz, J. (2009). Optimization of the continuous biosorption of copper with sugar-beet pectin gels. *Journal of Environmental Management*, 90(5), 1737-1743.
41. Mathew, B., & Krishnamurthy, N. (2018). Screening and identification of bacteria isolated from industrial area groundwater to study lead sorption: Kinetics and statistical optimization of biosorption parameters. *Groundwater for Sustainable Development*, 7, 313-327.
42. Moreira, V., Lebron, Y., Freire, S., Santos, L., Palladino, F., & Jacob, R. (2019). Biosorption of copper ions from aqueous solution using *Chlorella pyrenoidosa*: Optimization, equilibrium and kinetics studies. *Microchemical Journal*, 145, 119-129.
43. Morosanu, I., Teodosiu, C., Coroaba, A., & Paduraru, C. (2019). Sequencing batch biosorption of micropollutants from aqueous effluents by rapeseed waste: Experimental assessment and statistical modelling. *Journal of Environmental Management*, 230, 110-118.
44. Mukhopadhyay, M., Noronha, S., & Suraishkumar, G. (2007). Kinetic modeling for the biosorption of copper by pretreated *Aspergillus niger* biomass. *Bioresource Technology*, 98(9), 1781-1787.
45. Mukhopadhyay, M., Noronha, S., & Suraishkumar, G. (2008). Copper biosorption in a column of pretreated *Aspergillus niger* biomass. *Chemical Engineering Journal*, 144(3), 386-390.
46. Nakajima, A. (2002). Electron spin resonance study of copper biosorption by bacteria. *Water Research*, 36(8), 2091-2097.
47. Nazari, A., Cox, P., & Waters, K. (2014). Biosorption of copper, nickel and cobalt ions from dilute solutions using BSA-coated air bubbles. *Journal of Water Process Engineering*, 3, 10-17.
48. Ngah, W., & Hanafiah, M. (2008). Biosorption of copper ions from dilute aqueous



- solutions on base treated rubber (Hevea brasiliensis) leaves powder: kinetics, isotherm, and biosorption mechanisms. *Journal of Environmental Sciences*, 20(10), 1168-1176.
49. Nishikawa, E., da Silva, M., & Vieira, M. (2018). Cadmium biosorption by alginate extraction waste and process overview in Life Cycle Assessment context. *Journal of Cleaner Production*, 178, 166-175.
  50. Ofomaja, A., Naidoo, E., & Modise, S. (2010). Biosorption of copper(II) and lead(II) onto potassium hydroxide treated pine cone powder. *Journal of Environmental Management*, 91(8), 1674-1685.
  51. Ofomaja, A., Naidoo, E., & Modise, S. (2010). Dynamic studies and pseudo-second order modeling of copper(II) biosorption onto pine cone powder. *Desalination*, 251(1), 112-122.
  52. Özdemir, U., Özbay, B., Özbay, İ., & Veli, S. (2014). Application of Taguchi L32 orthogonal array design to optimize copper biosorption by using Spaghnum moss. *Ecotoxicology and Environmental Safety*, 107, 229-235.
  53. Özer, A., Gürbüz, G., Çalimli, A., & Körbahti, B. (2009). Biosorption of copper(II) ions on Enteromorpha prolifera: Application of response surface methodology (RSM). *Chemical Engineering Journal*, 146(3), 377-387.
  54. Padilha, F., de França, F., & da Costa, A. (2005). The use of waste biomass of Sargassum sp. for the biosorption of copper from simulated semiconductor effluents. *Bioresource Technology*, 96(13), 1511-1517.
  55. Peng, H., Li, D., Ye, J., Xu, H., Xie, W., Zhang, Y., . . . Liu, W. (2019). Biosorption behavior of the Ochrobactrum MT180101 on ionic copper and chelate copper. *Journal of Environmental Management*, 235, 224-230.
  56. Peng, S.-H., Wang, R., Yang, L.-Z., He, L., He, X., & Liu, X. (2018). Biosorption of copper, zinc, cadmium and chromium ions from aqueous solution by natural foxtail millet shell. *Ecotoxicology and Environmental Safety*, 165, 61-69.
  57. Podstawczyk, D., Witek-Krowiak, A., Dawiec, A., & Bhatnagar, A. (2015). Biosorption of copper(II) ions by flax meal: Empirical modeling and process optimization by response surface methodology (RSM) and artificial neural network (ANN) simulation. *Ecological Engineering*, 83, 364-379.

58. Preetha, B., & Viruthagiri, T. (2007). Application of response surface methodology for the biosorption of copper using *Rhizopus arrhizus*. *Journal of Hazardous Materials*, 143(1), 506-510.
59. Pundir, R., Chary, G., & Dastidar, M. (2018). Application of Taguchi method for optimizing the process parameters for the removal of copper and nickel by growing *Aspergillus* sp. *Water Resources and Industry*, 20, 83-92.
60. Puyen, Z., Villagrasa, E., Maldonado, J., Diestra, E., Esteve, I., & Solé, A. (2012). Biosorption of lead and copper by heavy-metal tolerant *Micrococcus luteus* DE2008. *Bioresource Technology*, 126, 233-237.
61. Ronda, A., Martín-Lara, M., Dionisio, E., Blázquez, G., & Calero, M. (2013). Effect of lead in biosorption of copper by almond shell. *Journal of the Taiwan Institute of Chemical Engineers*, 44(3), 466-473.
62. Sahmoune, M. (2018). Performance of *Streptomyces rimosus* biomass in biosorption of heavy metals from aqueous solutions. *Microchemical Journal*, 141, 87-95.
63. Schut, S., Zauner, S., Hampel, G., König, H., & Claus, H. (2011). Biosorption of copper by wine-relevant lactobacilli. *International Journal of Food Microbiology*, 145(1), 126-131.
64. Sedlakova-Kadukova, J., Kopcakova, A., Gresakova, L., Godany, A., & Pristas, P. (2019). Bioaccumulation and biosorption of zinc by a novel *Streptomyces* K11 strain isolated from highly alkaline aluminium brown mud disposal site. *Ecotoxicology and Environmental Safety*, 167, 204-211.
65. Sheel, A., & Pant, D. (2018). Recovery of gold from electronic waste using chemical assisted microbial biosorption (hybrid) technique. *Bioresource Technology*, 247, 1189-1192.
66. Sheng, P., Wee, K., Ting, Y., & Chen, J. (2008). Biosorption of copper by immobilized marine algal biomass. *Chemical Engineering Journal*, 136(2), 156-163.
67. Terry, P., & Stone, W. (2002). Biosorption of cadmium and copper contaminated water by *Scenedesmus abundans*. *Chemosphere*, 47(3), 249-255.
68. Trinelli, M., Areco, M., & dos Santos Afonso, M. (2013). Co-biosorption of copper and glyphosate by *Ulva lactuca*. *Colloids and Surfaces B: Biointerfaces*, 105, 251-258.

69. Uçun, H., Aksakal, O., & Yildiz, E. (2009). Copper(II) and zinc(II) biosorption on *Pinus sylvestris* L. *Journal of Hazardous Materials*, 161(2), 1040-1045.
70. Vieira, L., de Araujo, L., de Padua Ferreira, R., da Silva, E., Canevesi, R., & Marumo, J. (2019). Uranium biosorption by *Lemna* sp. and *Pistia stratiotes*. *Journal of Environmental Radioactivity*, 203, 179-186.
71. Vishan, I., Saha, B., Sivaprakasam, S., & Kalamdhad, A. (2019). Evaluation of Cd(II) biosorption in aqueous solution by using lyophilized biomass of novel bacterial strain *Bacillus badius* AK: Biosorption kinetics, thermodynamics and mechanism. *Environmental Technology & Innovation*, 14, 100323.
72. Wan Maznah, W., Al-Fawwaz, A., & Surif, M. (2012). Biosorption of copper and zinc by immobilised and free algal biomass, and the effects of metal biosorption on the growth and cellular structure of *Chlorella* sp. and *Chlamydomonas* sp. isolated from rivers in Penang, Malaysia. *Journal of Environmental Sciences*, 24(8), 1386-1393.
73. Wang, J.-Y., Cui, H., Cui, C.-W., & Xing, D.-F. (2016). Biosorption of copper(II) from aqueous solutions by *Aspergillus niger*-treated rice straw. *Ecological Engineering*, 95, 793-799.
74. Wang, L., Liu, X., Lee, D.-J., Tay, J.-H., Zhang, Y., Wan, C.-L., & Chen, X.-F. (2018). Recent advances on biosorption by aerobic granular sludge. *Journal of Hazardous Materials*, 357, 253-270.
75. Xie, Y.-d., Xiong, W.-l., Yu, J.-x., Tang, J.-Q., & Chi, R.-a. (2018). Recovery of copper from metallurgical sludge by combined method of acid leaching and biosorption. *Process Safety and Environmental Protection*, 116, 340-346.
76. Xu, H., Tan, L., Cui, H., Xu, M., Xiao, Y., Wu, H., . . . Xie, J. (2018). Characterization of Pd(II) biosorption in aqueous solution by *Shewanella oneidensis* MR-1. *Journal of Molecular Liquids*, 255, 333-340.
77. Yahaya, Y., Mat Don, M., & Bhatia, S. (2009). Biosorption of copper (II) onto immobilized cells of *Pycnopus sanguineus* from aqueous solution: Equilibrium and kinetic studies. *Journal of Hazardous Materials*, 161(1), 189-195.
78. Yargıç, A., Yarbay Şahin, R., Özbay, N., & Önal, E. (2015). Assessment of toxic



copper(II) biosorption from aqueous solution by chemically-treated tomato waste. *Journal of Cleaner Production*, 88, 152-159.

79. Klich, M. A. (2002). *Identification of common Aspergillus species*. Centraalbureau voor Schimmelcultures (CBS).
80. Moss, S. T., & Cheadle, M. A. (2000). The fungal genus *Aspergillus*: A review of its cultural and morphological characteristics. *Mycological Research*, 104(3), 318-325.  
<https://doi.org/10.1017/S0953756299002549>
81. Volesky, B. (2007). Biosorption and bioaccumulation of heavy metals. In *Handbook of Environmental Chemistry* (pp. 1-38). Springer, Berlin, Heidelberg.  
[https://doi.org/10.1007/978-3-540-47609-9\\_1](https://doi.org/10.1007/978-3-540-47609-9_1)
82. Aksu, Z., & Tezer, S. (2005). Biosorption of reactive dyes by dried *Aspergillus niger*: A comparative study. *Journal of Hazardous Materials*, 122(1-2), 7-14.  
<https://doi.org/10.1016/j.jhazmat.2005.01.030>
83. Pino, I., et al. (2006). Effect of agitation on cadmium and copper biosorption by *Aspergillus niger* from aqueous solutions. *Journal of Environmental Management*, 81(2), 144-150. <https://doi.org/10.1016/j.jenvman.2006.06.021>
84. Anand, N., Muthukumar, K., & Srinivasan, P. (2020). Optimization of biosorption for the removal of heavy metals using *Aspergillus* sp. *Environmental Science and Pollution Research*, 27(3), 3409-3419. <https://doi.org/10.1007/s11356-019-07075-0>
85. Meena, S. M., Baskar, S., & Srinivasan, P. (2008). Biosorption of heavy metals by fungal biomass. *Bioresource Technology*, 99(9), 3056-3061.  
<https://doi.org/10.1016/j.biortech.2007.08.051>



# SWARNIM

## STARTUP & INNOVATION UNIVERSITY

WHERE IDEAS COME ALIVE

Date: 13/12/2021

### Statement of Expenditure/Utilization certificate

This is to certify that the seed money sanctioned to **Dr. Priti Mahla, Assistant Professor, Swarnnim Science College** in the academic year 2021-22 has been utilized as per the following details:

Title of Project	Amount Sanctioned in (Rs)	Amount disbursed in (Rs)		Actual Expenditure in (Rs)
		1-6 months	6-12 months	
Copper biosorption utilizing dehydrated Aspergillus biomass	500000	309000	50000	359000



Accounts/Finance officer

Swarnnim Startup and Innovation University